Genome Size Survey of the Japanese Pine Sawyer Beetle, *Monochamus alternatus* (Coleoptera: Cerambycidae)

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**Authors’ contributions**

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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**ABSTRACT**

*Monochamus alternatus* is a longicorn beetle, an important stem borer of pine trees in China. In this study, we determined the first accurate genomic survey of *M. alternatus* by the next-generation sequencing (NGS), which is a relatively new methodology that can ensure the identification of large numbers of simple sequences repeat (SSR) markers, in order to increases the abundance of the Cerambycidae genome information and assist in phylogenetic, molecular systematics and evolutionary studies for Coleoptera. The result showed that the genome size was 871.09 Mb, the GC content of the genome was within normal limits (34.45%), the proportion of repetitive sequences was high (59.40%), and the heterozygosity rate was low (1.04%). The heterozygosity of *M. alternatus* is higher than 0.5% and the repeat rate is more than 50%. Based on this we inferred the use of Illumina+PacBio in sequencing assembly strategy is highly recommended.

Keywords: Coleoptera; Monochamus alternates; genome survey; high-throughput sequencing.

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1. INTRODUCTION

*Monochamus alternatus*, belonging to the subfamily Lamiinae of Coleoptera, is not only an important stem borer of pine trees in China, but also the main vector of pine wood nematode disease [1]. *M. alternatus* plays a key role in carrying, spreading and assisting the pathogen to invade the host in the process of spread and infection of pine wood nematode [2]. Natural enemies include pathogenic microorganisms, parasitic nematodes, parasitic insects, predatory insects, spiders, birds and so on. The life cycle can be divided into four stages: pupa, larva, egg and adult, with one generation a year, and the mature larva overwinters in the tunnel. Distributed in China, South Korea, Japan, Laos. It was listed on the list of forestry dangerous pests of the State Forestry Administration of China in 2003 [3]. In terms of its genetic characteristics, the whole genome of *M. alternatus* has not yet been published [4,5]. In view of the great harm of this species to forestry, we use the next-generation sequencing (NGS) technology to determine and analyze its whole genome in order to provide basic molecular data for further study of its behavior, physiology, ecology, genetic variation, evolutionary adaptation, population protection and the formulation of conservation strategies [6-11].

Despite its great harmless to pine wood, the genetic information of *M. alternatus* has remained largely unknown, the genome size and genome-wide sequencing of *M. alternatus* have not been reported. The mitogenomes of *M. alternatus*, *Bursaphelenchus mucronatus*, and some of the closely related species have been studied [12]. Formerly, scholars have used morphology, anatomy and physiology, and other methods to establish relationships among species for creating classification systems. However, the evolutionary relationships between *M. alternatus* and other closely related species are hard to describe just based on traditional dates. Previous studies have used mitogenome dates to assess the gene expression of *M. alternatus* [13]. Thus, we aim to expand the database of Lamiinae and provide some useful information for further study.

With the development of the sequencing techniques, more and more manuscript genomes have been assembled. The main difference between genomics and traditional biological methods lies in the scale of research, because the goal of genomics is to analyze a large number of genes extensively and may even involve a complete set of genes that make up the genome, rather than being limited to one or a small number of genes. With the development of DNA sequence technology, the cost has been decreased rapidly and the ability of obtaining the date has been increased vastly. NGS technology has significantly increased the sequence output while reducing time and cost. Genomic survey is a method that combine NGS technology with K-mer dates to achieve species genome size, GC content, heterozygosity rate and repetition rate. This technology has been used widely and could also accurately predict the whole genome sizes.

In this study, we provided valuable molecular data to reveal the relationship between *M. alternatus* and pine wood nematode [14] proved that *M. alternatus* infected with *B. xylophilus* showed an increase in the expression of some antioxidant genes, so that it could obtain immune tolerance, this study showed that the existence of *B. xylophilus* would increase the expression level of some metabolic, so some scholars inferred that the relationship between nematodes and beetles should be host switching rather than nematode-beetle coevolution, because in their study, they found some efforts were made by *M. alternatus* to offset the impact. The mechanisms need to be studied in the future.

2. METHODS AND MATERIALS

During the inspection in An’xi county of Quanzhou, we had obtained the *M. alternatus* specimens, which were frozen in liquid nitrogen and stored in a cryogenic refrigerator at 80 degrees below zero.

The genomic DNA, was extracted by improved SDS method. The Absorbance values of 260 nm and 280 nm were determined by ultraviolet spectrophotometer to estimate the concentration and purity of the extracted DNA, and then agarose gel electrophoresis was performed to detect the integrity of the extracted DNA. When sampling, 1 kb DNA Ladder (Takara) and λ-Hind di-gest (Takara) were used as Marker, gel concentration of 0.8%, electrophoresis for 60 minutes under the condition of 80V voltage. After the electrophoresis was completed, the electrophoretic band of genomic DNA was observed in the gel imaging system of BIO-RAD company.

The DNA samples were randomly interrupted by Covaris ultrasonic crusher, and the small
fragment sequencing library of 250 bp was established. Then the whole library was prepared by terminal repair, a tail, sequencing joint, purification, PCR amplification and so on. After that, double-terminal (Pair-End) sequencing was carried out on the Illumina HiseqTM2000 platform. Finally, the quality control is carried out by using the software NGS-QC-Generator, the low-quality data is filtered out, and the effective data obtained are used for genome feature evaluation and preliminary assembly.

K-mer analysis was used to estimate the genome size. Use the formula: genome size = K-mer number / peak depth to estimate genome size. The estimated value of K-mer depth is calculated, which is used to estimate the genome size. The heterozygosity (Φ) was calculated by the following formula, while the repetition rate was calculated by calculating the ratio of the number of K-mer to the total number after 1.8 times the depth of the homozygous peak. In the formula, K is the number of heterozygous K-mer, \( a_{1/2} \) is the percentage of heterozygous K-mer species, and \( n_{Kspecies} \) is the number of all K-mer species.

The GC depth content of the target genome was calculated by constructing the GC dot map from the total number of bases in the sequencing data, and the correlation analysis was carried out. Contigs were rearranged by all clean reads and scaffolds were gradually constructed by diversified insert size paired-ends.

The sequence reads obtained from all the small fragment libraries were truncated into smaller sequence fragments, and the de-Brujin map was constructed by using the overlapping relationship between them. Then the simplified de-Brujin map which is easy to analyze was obtained by screening branches, simplifying the bifurcation pathway and randomly combining heterozygous sites, and then the bifurcation sequence was truncated into small fragments to get the initial overlapping (contigs). The reads obtained by sequencing was compared, and the obtained contigs, assembled the contigs into scaffolds by using the connection relationship between reads and the size information of insertion fragments. In order to make the assembled sequence more complete, it is necessary to connect the contigs, according to the pairing relationship between the double-terminal data and optimize and fill holes in the gaps between the contig, so as to obtain the original genome sequence.

3. RESULTS

3.1 Sequencing Quality

The low-quality data is filtered out by screening, and the effective data with the size of 34.68Gb is obtained, and the sequencing depth is 51x. Through the strict filtering of the obtained data, the high-quality pure data is obtained. The 350 bp library is constructed, the original data is 34.683,801,900bp, and the clean data 34,652,586,478bp is obtained after filtering. Both Q20 and Q30 are indicators to measure the quality of sequencing. It is generally believed that when Q20 ≥ 90% and Q30 ≥ 80%, the quality of sequencing data is better. In this study, the content of Q20 is 97.86% and the content of Q30 is 93.48%. It is known that the sequencing quality of this study is good, and the sequencing error rate is 0.04%, which is also within the normal range (< 0.05%).

After genome regulation, if the estimated heterozygosity of the genome is high (higher than 0.5%), or the content of repetitive sequences is high (more than 50%), the genome can be regarded as a complex genome. WGS sequencing and assembly strategies suitable for general genomes are generally difficult to obtain good complex genome maps. At present, there are two commonly used strategies to solve complex genome maps. SangeH-454 sequencing and WGS+BAC to BAC/fosmid to fosmid. Compared with the former, the latter is based on Illumina sequencing, and the cost is much lower. In this study, we found the heterozygosity of M. alternatus is higher than 0.5%, the repeat rate is more than 50% either. In consequence, the use of Illumina+PacBio in sequencing assembly strategy is highly recommended.

3.2 17-Mer Analysis and Genome Size Estimation

Genome size of M. alternatus was estimated according to the Lander waterman algorithm based on K-mer (k = 17) frequency of the clean reads and the 17-mer frequency distribution complied with the poisson distribution. The 34.65 Gb valid data of M. alternatus were analyzed by K-mer (Fig. 1). It can be seen that the peak is near depth = 29; the calculated genome size is 871.09 Mbp, the modified genome size is 860.06 Mbp. There is a tail on the K-mer curve, indicating that the proportion of genome repeat sequences of M. alternatus is large.
Fig. 1. 17 K-mer analysis for estimating the genome size of Monochamus alternatus

Note: The genome size was estimated by using the formula: Genome size = \( K \)-me num/Peak depth

Table 1. Statistics of assembly results in the Monochamus alternatus genome

<table>
<thead>
<tr>
<th>Item</th>
<th>Contig</th>
<th>Scaffold</th>
</tr>
</thead>
<tbody>
<tr>
<td>N50</td>
<td>787</td>
<td>954</td>
</tr>
<tr>
<td>N90</td>
<td>135</td>
<td>145</td>
</tr>
<tr>
<td>Total Size</td>
<td>823,399,058</td>
<td>840,047,447</td>
</tr>
<tr>
<td>Total Number</td>
<td>2,034,890</td>
<td>1,839,509</td>
</tr>
<tr>
<td>Longest</td>
<td>385,511</td>
<td>458,244</td>
</tr>
</tbody>
</table>

3.3 Results of Preliminary Assembly of the Genome

Using SOAP-Denovo software to assemble the data, the initial genome sequence is obtained, and the splicing result is shown in Table 1. It can be seen that the genomic Contig N50 of *M. alternatus* is 787 bp and scaffolding N50 is 954 bp, which is short, so it is not suitable for further assembly by shotgun method.

3.4 Content and Distribution of GC

GC content is an important character of nucleic acid sequence composition in living creatures, which is also used to detect the separation of AT and GC. Theoretically, the contents of G and C bases and the A and T bases should be equal in each sequencing cycle, and the whole sequencing process is stable and horizontal. In the actual sequencing process, due to the DNA template amplification deviation and the low sequencing quality value of the first few bases, it often leads to large fluctuations in the first few bases of each read, which is a normal situation. According to the statistics of GC content of assembled contig, the distribution map of GC content and sequencing depth (depth) was obtained (Fig. 2). The genomic GC content of *M. alternatus* was 32.94%.

3.5 Genome Comparison of the Insects in Coleoptera

In the genome database of the National Biotechnology Information Center of the United States (https://www.ncbi.nlm.nih.govgenome), enter the key words “Coleoptera” to obtain 61 pieces of genome information of Coleoptera insects. We select 25 species that can be reported in the relevant literatures, and compare their genome information with the genome data of *M. alternatus* (Table 2). According to the published data, the genome sizes of Cerambycidae range from 90.87 to 1112.44 Mb, while the genome size of *M. alternatus* is as high as 871.09 Mb. Some studies believed that if the content of genomic GC is moderate (25% < GC < 65%), it will not affect the accuracy of genome sequencing and correct assembly. The content of genomic GC measured in this study is 32.94%, so the sequencing results and assembly should be correct. On the other hand, the GC contents of the other 24 genomes were similar.
4. DISCUSSION

Although the depth of our *M. alternatus* preliminary sequencing genome is relatively low and it's a disadvantage for an acquaintance of genome information, we still excavated some basic genome information and developed genome resources by using several bioinformation methods like K-mer and scaffold assembly methods. According to each analysis index, it is inferred that the genome of *M. alternatus* is a highly complex genome, and the corresponding strategy can be used to assemble the genome.

There are many species of insects, about 1.8 million species are known, accounting for more than 3 beat 4 of the described animal species in the world [15-18]. In recent years, with the continuous improvement of DNA sequencing technology in the direction of automation [19], generalization and low cost, the continuous maturity of genomics research technology and the continuous updating of bioinformatics analysis methods [20], many entomologists use various ensemble research methods such as genome, transcriptome, proteome, etc. to obtain a large number of molecular biological data, and use bioinformatics methods to analyze and mine the data. This paper attempts to reveal the genetic basis of the special physiological activities and behaviors of some insects from the point of view of molecular biology [21,22], expounding the molecular mechanism of co-evolution of insects and other organisms [23], showing a broad research prospect of insect molecular biology [24,25].

Coleoptera is the first order with the largest number of species and the widest distribution in the Insecta and even the animal kingdom. There are many kinds and the system is complex. However, there are also many species (mainly larvae) that harm crops and bring loss or inconvenience to human production and life [26]. Therefore, the research on Coleoptera insect genomics has important theoretical significance and application value. Nowadays, the genomic research of Coleoptera insects is mainly to analyze their special habits or behaviors at the genome level [27-30]. Genome size, also known as C-value or Constant-value, is an important

![Fig. 2. GC content and average sequencing depth](image-url)
Fig. 3. 17 K-mer analysis for estimating the genome size of Monochamus alternatus

genetic feature of an organism. M. alternatus is known as the cancer of pine trees and is also a devastating disease, causing serious environmental and economic losses. Effective strategies are needed to stop or control the spreading of this disease. This study will provide a method to assemble the genome and may serve as key point to develop new control strategies for pine wilt disease, the results of the current study will play an important role in future whole-genome sequencing projects and provide an abundant resource for further functional studies, which will help us learn genetic regulatory mechanisms.

Table 2. Comparison of the mitogenome assembly data of Monochamus alternatus with that of 25 species of Coleoptera

<table>
<thead>
<tr>
<th>Species</th>
<th>Genome size (Mb)</th>
<th>Number of protein-coding genes</th>
<th>GC content (%)</th>
<th>Heterozygosity (%)</th>
<th>Repeat (%)</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Monochamus alternatus</td>
<td>871.09</td>
<td>n.a.</td>
<td>32.94</td>
<td>1.04</td>
<td>59.04</td>
<td>This study</td>
</tr>
<tr>
<td>Diabrotica virgifera</td>
<td>2418.07</td>
<td>28061</td>
<td>36.5</td>
<td>n.a.</td>
<td>n.a.</td>
<td>[15]</td>
</tr>
<tr>
<td>Photinus pyralis</td>
<td>471.51</td>
<td>32294</td>
<td>36.4</td>
<td>0.598</td>
<td>42.6</td>
<td>[31]</td>
</tr>
<tr>
<td>Listronotus bonariensis</td>
<td>1112.44</td>
<td>n.a.</td>
<td>31.3</td>
<td>0.18</td>
<td>70</td>
<td>[30]</td>
</tr>
<tr>
<td>Propylea japonica</td>
<td>851.23</td>
<td>18018</td>
<td>35.13</td>
<td>0.9</td>
<td>58.22</td>
<td>[32]</td>
</tr>
<tr>
<td>Marronius borbonicus</td>
<td>406.94</td>
<td>23278</td>
<td>35.9</td>
<td>0.2</td>
<td>29.2</td>
<td>[33]</td>
</tr>
<tr>
<td>Abscondita terminalis</td>
<td>499.65</td>
<td>20439</td>
<td>31.4</td>
<td>n.a.</td>
<td>n.a.</td>
<td>[34]</td>
</tr>
<tr>
<td>Lamprigera yunnana</td>
<td>1052.93</td>
<td>19438</td>
<td>34.1</td>
<td>n.a.</td>
<td>n.a.</td>
<td>[34]</td>
</tr>
<tr>
<td>Hycleus cichorii</td>
<td>99.17</td>
<td>13813</td>
<td>32.3</td>
<td>1.00</td>
<td>72.73</td>
<td>[35]</td>
</tr>
<tr>
<td>Species</td>
<td>Genome size (Mb)</td>
<td>Number of protein-coding genes</td>
<td>GC content (%)</td>
<td>Heterozygosity (%)</td>
<td>Repeat (%)</td>
<td>References</td>
</tr>
<tr>
<td>-------------------------</td>
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<td>--------------------------------</td>
<td>----------------</td>
<td>-------------------</td>
<td>------------</td>
<td>------------</td>
</tr>
<tr>
<td>Harmonia axyridis</td>
<td>466.692</td>
<td>n.a.</td>
<td>33.2</td>
<td>n.a.</td>
<td>n.a.</td>
<td>[36]</td>
</tr>
<tr>
<td>Hycleus phaleratus</td>
<td>90.87</td>
<td>13725</td>
<td>30.06</td>
<td>0.99</td>
<td>74.90</td>
<td>[35]</td>
</tr>
<tr>
<td>Aethina tumida</td>
<td>234.34</td>
<td>17463</td>
<td>30.00</td>
<td>n.a.</td>
<td>n.a.</td>
<td>[37]</td>
</tr>
<tr>
<td>Limonius californicus</td>
<td>1072.67</td>
<td>n.a.</td>
<td>35.00</td>
<td>0.21</td>
<td>n.a.</td>
<td>[38]</td>
</tr>
<tr>
<td>Oryctes borbonicus</td>
<td>406.20</td>
<td>8822</td>
<td>34.85</td>
<td>0.2</td>
<td>29.2</td>
<td>[33]</td>
</tr>
<tr>
<td>Nicrophorus vespilloides</td>
<td>195.27</td>
<td>18995</td>
<td>32.20</td>
<td>n.a.</td>
<td>n.a.</td>
<td>[22]</td>
</tr>
<tr>
<td>Asbolus verrucosus</td>
<td>249.61</td>
<td>n.a.</td>
<td>32.90</td>
<td>n.a.</td>
<td>n.a.</td>
<td>[39]</td>
</tr>
<tr>
<td>Protaetia brevivarsis</td>
<td>1143.92</td>
<td>n.a.</td>
<td>25.40</td>
<td>72.29</td>
<td>n.a.</td>
<td>[40]</td>
</tr>
<tr>
<td>Popilia japonica</td>
<td>531.53</td>
<td>n.a.</td>
<td>34.90</td>
<td>n.a.</td>
<td>n.a.</td>
<td>[41]</td>
</tr>
<tr>
<td>Anoplophora glabripennis</td>
<td>706.97</td>
<td>20632</td>
<td>33.40</td>
<td>n.a.</td>
<td>n.a.</td>
<td>[25]</td>
</tr>
<tr>
<td>Tenebrio molitor</td>
<td>280.78</td>
<td>n.a.</td>
<td>26.50</td>
<td>n.a.</td>
<td>n.a.</td>
<td>[42]</td>
</tr>
<tr>
<td>Agrilus planipennis</td>
<td>353.07</td>
<td>22159</td>
<td>36.00</td>
<td>n.a.</td>
<td>n.a.</td>
<td>[17]</td>
</tr>
<tr>
<td>Leptinotarsa decemlineata</td>
<td>641.99</td>
<td>19038</td>
<td>35.60</td>
<td>n.a.</td>
<td>n.a.</td>
<td>[43]</td>
</tr>
<tr>
<td>Callosobruchus maculatus</td>
<td>1007.82</td>
<td>31345</td>
<td>37.7</td>
<td>n.a.</td>
<td>65.00</td>
<td>[44]</td>
</tr>
<tr>
<td>Priacma serrata</td>
<td>12.08</td>
<td>n.a.</td>
<td>35.90</td>
<td>n.a.</td>
<td>n.a.</td>
<td>[45]</td>
</tr>
<tr>
<td>Dendroctonus ponderosae</td>
<td>257.09</td>
<td>16791</td>
<td>38.45</td>
<td>n.a.</td>
<td>n.a.</td>
<td>[33]</td>
</tr>
</tbody>
</table>

Fig. 4. 17 K-mer analysis for estimating the genome size of *Monochamus alternatus*
This study lays the foundation of whole-genome sequencing in *M. alternatus*, and puts forward the platform for the next investigation on this particular insect. Today, more and more scholars studied the relationship between *M. alternatus* and pine wood nematode, because of its great harm to the pine wood. Our study increases the abundance of the Cerambycidae genome information and can assist in phylogenetic, molecular systematics and evolutionary studies of Cerambycidae. However, molecular data on species of Lamiinae are still scarce and more information is needed to fully explore the relationships within Lamiinae.

5. CONCLUSION

The sequencing quality of this study was good and within the normal range. The K-mer data indicated that there is a large proportion of repeats in the genome of *M. alternatus*. The genome GC content is moderate, which will not affect the accuracy and correct assembly of genome sequencing. The genome of *M. alternatus* is a complex genome with high repeat sequences.

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES


